
Euferol and Melliferol: Two Novel Triterpenoids from *Euphorbia mellifera*

M. José U. Ferreira

Faculty of Pharmacy, University of Lisbon, Av. das Forças Armadas, 1600 Lisbon, Portugal

Ana M. Lobo

Centro de Química Estrutural, Complexo I, INIC, Av. Rovisco Pais, 1096 Lisbon, and Secção de Química Orgânica Aplicada, FCT, New University of Lisbon, Quinta da Torre, 2825 Monte de Caparica, Portugal

Caroline A. O'Mahoney and David J. Williams

Department of Chemistry, Imperial College, London SW7 2AY, U.K.

Hugo Wyler

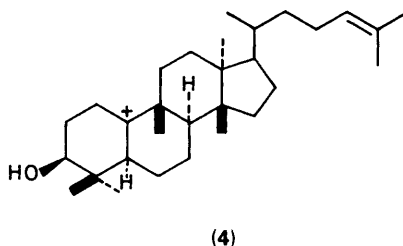
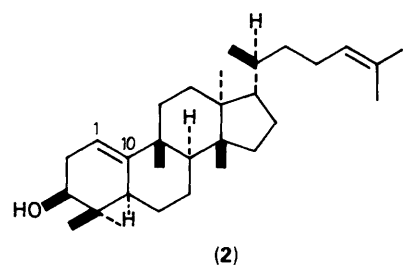
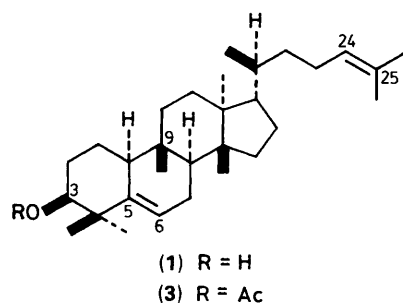
Institut of Organic Chemistry, University of Lausanne, Rue de la Barre, 1005 Lausanne, Switzerland

The structures and stereochemistries of two novel skeletons euferol [19(10→9)*abeo*-8 α ,9 β ,10 α -tirucall-5-en-3 β -ol] and melliferol [19(10→9)*abeo*-5 α ,8 α ,9 β -tirucall-1(10)-en-3 β -ol], found in *Euphorbia mellifera*, have been established from spectral data and single crystal X-ray analyses.

Euphorbia mellifera Ait., an Euphorbiaceae commonly encountered in the Atlantic island of Madeira, has been found to contain two triterpenes euferol (1) and melliferol (2), corresponding respectively to the new skeletons 19(10→9)*abeo*-

8 α ,9 β ,10 α -tirucall-5-en-3 β -ol and 19(10→9)*abeo*-5 α ,8 α ,9 β -tirucall-1(10)-en-3 β -ol.

Euferol (1), C₃₀H₅₀O, by mass spectrometry, contained in its ¹H NMR spectrum (360 MHz, CDCl₃) five singlets attributed



to tertiary methyl groups (δ 0.81, 0.84, 0.87, 1.06, and 1.14), two methyl groups attached to an sp^2 carbon (δ 1.60 and 1.68), one secondary methyl group [δ 0.92 (d, J 6 Hz)], two olefinic protons at δ 5.11 and 5.67, and a proton attached to C(3) [δ 3.45 (t, J 2.5 Hz)] with features characteristic of equatorial configuration.¹ Euferyl which could not be induced to crystallize yielded however a crystalline monoacetate which gave essentially the same ^1H NMR spectrum as (1), except for the proton attached to C(3) which now appeared at δ 4.72 (t, J 2.5 Hz). Of the two olefinic protons, the one at δ 5.11 [the same as in (1)] was assigned to the C(24) hydrogen and the other which had suffered a small shift to δ 5.57, in relation to (1), was ascribed to the olefinic proton at C(6).¹ The retro-Diels–Alder reaction involving cleavage of ring B observed in the mass spectrum of (1) and (3) further pointed towards the presence of a double bond at C(5)–C(6).²

Melliferol (2), isomeric with (1), had similarly to (1), in its ^1H NMR spectrum (360 MHz, CDCl_3) five tertiary methyl groups (δ 0.64, 0.78, 0.84, 1.01, and 1.03), two methyls attached to a sp^2 carbon (δ 1.60 and 1.68), one secondary methyl [δ 0.91 (d, J 6 Hz)], two olefinic protons at δ 5.10 and 5.36, and a proton attached to carbon linked to an oxygen function [δ 3.52 (m, $w_{\frac{1}{2}}$ 20 Hz)].

The isomeric relationship between (1) and (2) became evident from a study of their respective ^{13}C NMR spectra. Thus signals in both spectra at δ 125.3 and 130.9 were attributed to the olefinic carbons at C(24) and C(25) respectively, and δ ca. 76 to the carbon bearing the alcohol function [C(3)]. Whereas the $\Delta^{5,6}$ carbons³ of (1) resonated at δ 142.1 and 121.9, the spectrum of (2) possessed a pair of signals, at δ 152.9 and 114.8 respectively, indicating that the olefinic bond is positioned

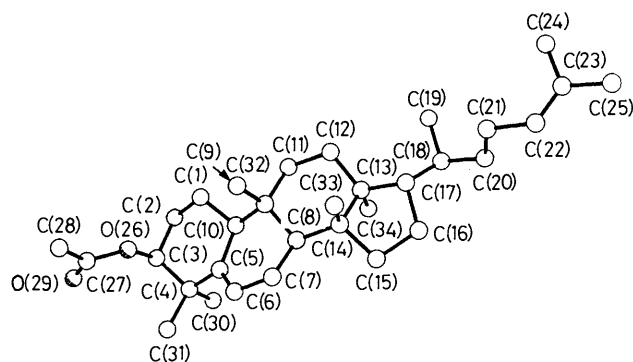


Figure 1. 3-D Structure of (3) obtained by X-ray analysis with atomic labelling. Oxygen atoms appear as shaded circles.

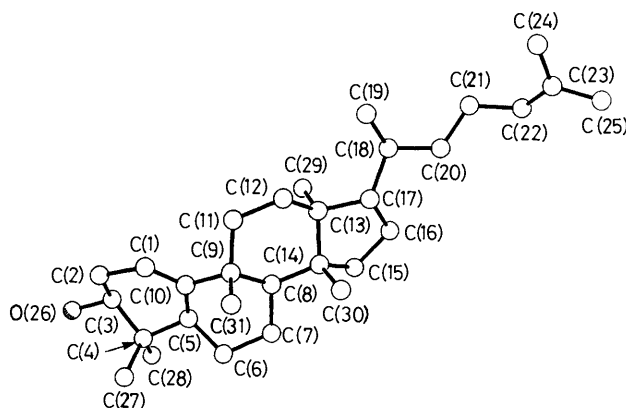


Figure 2. 3-D Structure of one of the pair of crystallographically independent molecules of (2) obtained by X-ray analysis with atomic labelling. Oxygen atoms appear as shaded circles.

differently in the triterpenoid nucleus. That it occupies the $\Delta^{1(10)}$ position (a very rare example) was suggested by the presence in its mass spectrum of two fragments m/z 354 and 72, resulting from a retro-Diels–Alder reaction, involving the cleavage of ring A.

The structure and stereochemistry of (2) and (3) were established unambiguously by X-ray analysis (Figures 1 and 2).^{*} These studies confirmed the C(1)–C(10) position for the double bond in (2) and its C(5)–C(6) sitting in (3). Also as deduced from the ^1H NMR data the C(3) proton in (3) is equatorial. In (2) this proton is axial.

It is interesting to speculate that if the carbocation (4) is indeed the biogenetic precursor of these two natural products,

* Crystal data.—For (2). $\text{C}_{30}\text{H}_{50}\text{O}$, $M = 426.7$, monoclinic, $a = 18.331(11)$, $b = 7.393(6)$, $c = 22.801(23)$ Å, $\beta = 111.47(6)^\circ$, $U = 2.876$ Å³, space group $P2_1$, $Z = 4$ (2 crystallographically independent molecules), $D_c = 0.99$ g cm⁻³, $\mu(\text{Cu-K}\alpha) = 4$ cm⁻¹. For (3). $\text{C}_{32}\text{H}_{52}\text{O}_2$, $M = 468.8$, monoclinic, $a = 12.391(4)$, $b = 7.452(2)$, $c = 15.626(6)$ Å, $\beta = 93.46(3)^\circ$, $U = 1.440$ Å³, space group $P2_1$, $Z = 2$, $D_c = 1.08$ g cm⁻³, $\mu(\text{Cu-K}\alpha) = 5$ cm⁻¹. Data for both structures were measured on a Nicolet R3m diffractometer with Cu-K α radiation (graphite monochromator) using ω -scans. Both structures were solved by direct methods, (2) was refined isotropically to give $R = 0.158$, $R_w = 0.158$ for 2316 independent observed reflections [$|F_o| \geq 3\sigma(|F_o|)$, $8 \leq 58^\circ$]; (3) was refined anisotropically to give $R = 0.047$, $R_w = 0.051$ for 1900 reflections. Isotropic only of (2) was performed because of the very poor quality and exceptionally small crystals available, and the consequently very limited data set. Atomic co-ordinates, bond lengths and angles, and thermal parameters have been deposited at the Cambridge Crystallographic Data Centre. See 'Notice to Authors', *J. Chem. Soc., Perkin Trans. 1*, 1989, Issue No. 1.

then their absolute stereochemistries would be depicted as in (1) and (2).

Experimental

Euferol (1) and Melliferol (2).—The oily fraction obtained from extraction of the aerial parts (3.2 kg) of *Euphorbia mellifera* with acetone and hexane followed by saponification (10% KOH, EtOH, 24 h, room temp.) was purified by argented column chromatography [SiO_2 , 20% AgNO_3 , eluting with hexane–EtOAc (95:5)] to yield a pure fraction of euferol (1) (350 mg) as a homogeneous colourless glass; m/z (%) 426 (M^+ , 2), 274 (11), 259 (19), 231 (5), 219 (4), 207 (29), 163 (15), 152 (1), and 134 (27). Acetylation of (1) (Ac_2O , py, 24 h, room temp.) gave white crystals of the monoacetate (3), m.p. 123–124 °C (Me_2CO); $[\alpha]_{\text{D}}^{25} +25.7$ (c 1.20, CHCl_3). Further elution yielded melliferol (2) (100 mg), m.p. 132–134 °C (Me_2CO), $[\alpha]_{\text{D}}^{25} -38.8$ °C (c 0.75, CHCl_3); m/z (%) 426 (M^+ ,

5), 354 (1), 259 (2), 241 (3), 231 (2), 219 (4), 207 (22), and 72 (50).

Acknowledgements

Calouste Gulbenkian Foundation is kindly thanked for a research grant (to M. J. U. F.) and Professor S. Prabhakar for helpful suggestions.

References

- 1 T. Itoy and T. Tamura, *Lipids*, 1980, **15**, 122.
- 2 C. B. Gamlath, A. A. L. Gunatilaka, and E. O. Schlemper, *J. Chem. Soc., Chem. Commun.*, 1988, 249.
- 3 J. B. Stothers, 'Carbon-13 NMR Spectroscopy,' Academic Press, New York, 1972, p. 441.

Paper 9/03961J

Received 6th June 1989

Accepted 18th September 1989